

## Preparation and NMR Structure of the Cyclo- $\beta$ -tripeptide [ $\beta^3$ -HGlu]<sub>3</sub> in Aqueous Solution: A New Class of Enterobactin-Type C<sub>3</sub>-Symmetrical Ligands?

Preliminary Communication

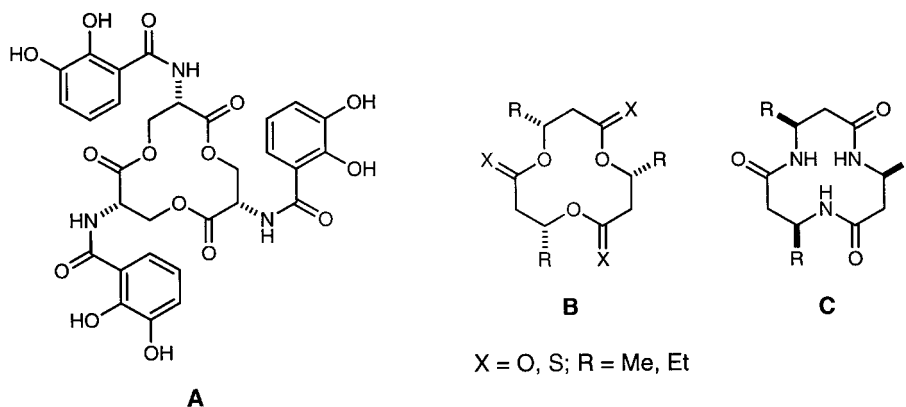
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Dedicated to Professor *Armin de Meijere*, Universität Göttingen, on the occasion of his 60th birthday

To date, cyclo- $\beta$ -tripeptides (twelve-membered-ring trilactams, **C**) have resisted structural investigations because of their extreme insolubility. Modelling and comparison with the corresponding tetramers indicate that the rings stack to form tube-like hydrogen-bonded aggregates (**D**). By exploiting the solubilizing effect of LiCl on peptides in THF, we were able to prepare the water-soluble title compound and determine its structure (**E**) by NMR spectroscopy. Structural similarities and differences between the trilactam ring of [ $\beta^3$ -HGlu]<sub>3</sub> and the corresponding trilactone ring, such as that of enterobactin (**A**), are discussed, and structures ('elongated'; **F**, **G**) are proposed that should be able to serve as tridentate or trivalent ligands for metal centers.

**Introduction.** – C<sub>3</sub>-Symmetrical cyclic oligomers such as *enterobactin* (**A**) play an important role in bacterial iron uptake and storage (*siderophores*; for a review article, see [1]); these compounds bind Fe<sup>III</sup> highly selectively and with extraordinary binding constants of up to 10<sup>49</sup> [2]. The X-ray structure of **A** with V<sup>IV</sup> revealed an all-up arrangement of the carbonyl groups with *s-trans*-conformation of the ester bonds and complexation of the metal ion by the catecholate units under the 'flat' macrocyclic ring [3].



<sup>1)</sup> Part of the projected dissertation of *K.G.*, ETH-Zürich

In the course of our work on the mechanism of the  $\text{Ca}^{2+}$  transport through phospholipid bilayers mediated by 3-hydroxybutanoic acid (HB) oligomers [4][5], we also prepared cyclic HB oligomers and found that the twelve-membered ring in the crystal structure of the trilactones **B** is almost superimposable with that of enterobactin [6–8]. The dipole moment of **B** ( $\text{X} = \text{O}$ ,  $\text{R} = \text{Me}$ ) was determined to be 4.6 D in  $\text{CCl}_4$  [7], consistent with the parallel arrangement of three  $\text{C}=\text{O}$  groups.

In contrast to the structures of the trilactones those of the corresponding highly insoluble cyclo- $\beta$ -tripeptides, such as **C**, are unknown<sup>2)</sup>. In the case of cyclo- $\beta$ -tetrapeptides consisting of  $\beta^3$ -HAla, we were able to determine (by powder X-ray diffraction) the structures of three diastereoisomers: they adopt stacked tubular structures with a tight net of H-bonds, so-called *peptide nanotubes* [10]<sup>3)</sup>. Another cyclo- $\beta$ -tetrapeptide has recently been shown to display biological activity mimicking that of a natural  $\alpha$ -peptide hormone [12].

**Modelling.** – In previous work on cyclo- $\beta$ -peptides with alkyl and with functionalized side chains [13–15], we had noticed the extremely poor solubility, and IR spectra indicated that these compounds also form a network of intermolecular H-bonds, maybe nanotube-type stacking in the solid state. Modelling of the parent compound (**C**;  $\text{R} = \text{H}$ ), with the MacroModel program [16], by placing seven trilactam units symmetrically on top of each other and allowing for energy minimization, indeed predicts the tube stacking as shown in the model **D** (*Scheme*). As can be seen, each of the rings is turned a few degrees against its neighbor (the angle  $\text{N}-\text{H}\cdots\text{O}$  deviates from  $180^\circ$ ). On each tetrahedral C-atom of the rings, there is an equatorial or lateral position available for substituents and an axial one occupied by H-atoms.

The infinite pleated-sheet-like stacking suggested by this model is, of course, compatible with the insolubility of such compounds in all common solvents except for  $\text{CF}_3\text{COOH}$ , and with their very high melting points of over  $200^\circ$  [14][15]. Four ways occurred to us to overcome such insolubility, *i.e.*, to break the intermolecular H-bonds and stabilize non-aggregated solubilized species: *i)* introduction of solubilizing side chains (bearing hydrophilic  $\text{NH}_3^+$  or  $\text{CO}_2^-$  groups), *ii)* complexation with ‘hard’ metal ions such as  $\text{Li}^+$ , which complex amide O-atoms very strongly [17]<sup>4)</sup>, *iii)* prohibition of stacking by switching to *N*-methyl-amide derivatives [17][19], and *iv)* prevention of intermolecular H-bonding by incorporating geminally disubstituted ( $\beta^{2,2}$  or  $\beta^{3,3}$ ) or, alternatively, *unlike*  $\beta^{2,3}$ -disubstituted  $\beta$ -amino-acid residues (these insure that a substituent is in a sterically forbidden position on a tetrahedral C-atom of the ring).

We have now made use of the first two methods, which allowed us to prepare and study the structure of a cyclo- $\beta$ -tripeptide in aqueous solution.

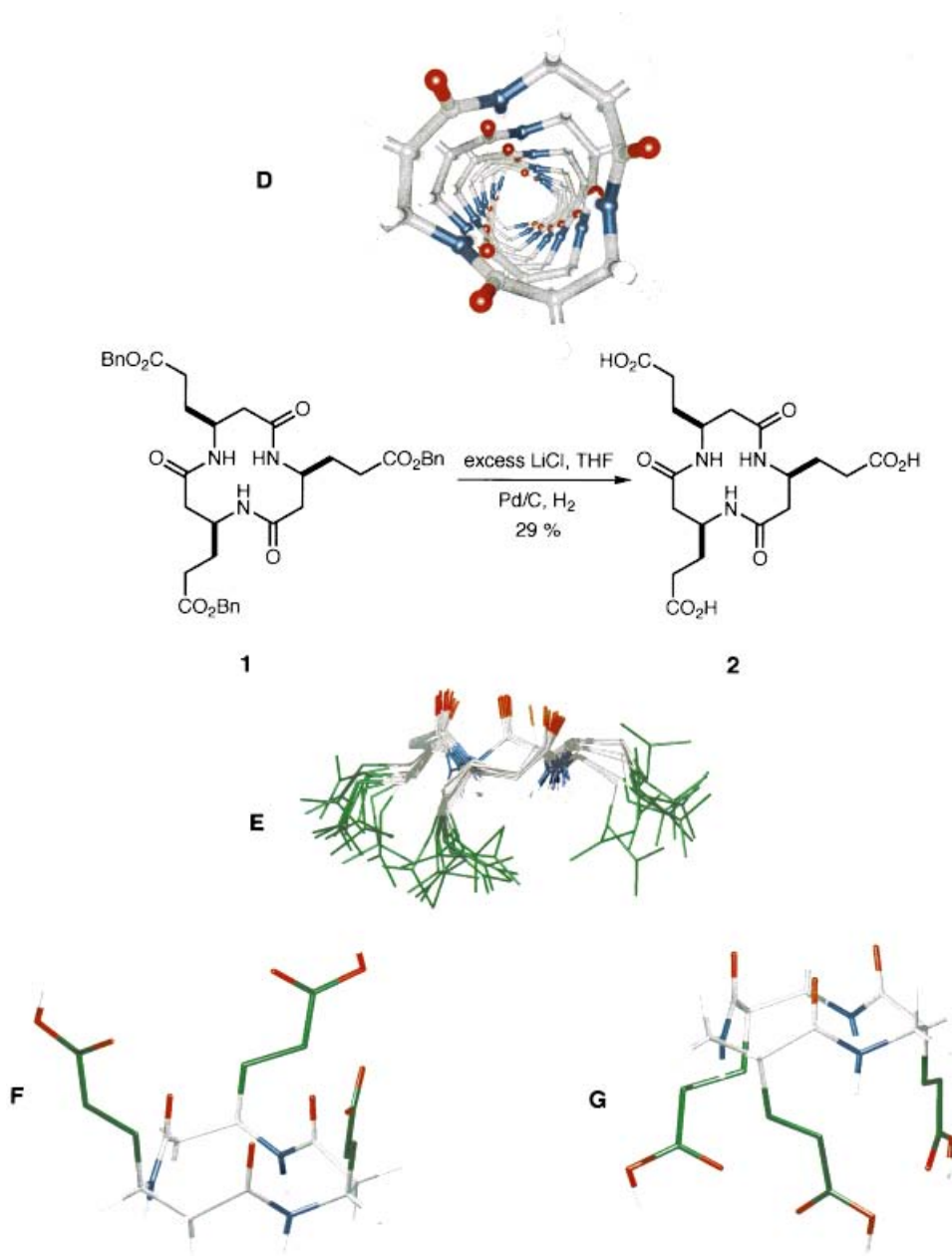
**Preparation of [ $\beta^3$ -HGlu]<sub>3</sub>.** – The trilactam **1** derived from glutamic acid with three benzyl-ester groups in the side chains, which could previously not be debenzylated for reasons of poor solubility [15], was dissolved in high concentrations in THF containing

<sup>2)</sup> Early molecular mechanics calculations by Drey and co-workers did not provide conclusive structural information about the achiral cyclo- $\beta$ -tripeptide **C** ( $\text{R} = \text{H}$ ) [9].

<sup>3)</sup> Incorporation of a cyclo- $\beta$ -tetrapeptide in phospholipid bilayers induces ion conductivity [11].

<sup>4)</sup>  $\Delta H^\circ$  values of up to  $-37$  kcal/mol have been measured [18].

Scheme. *Cyclo- $\beta$ -tripeptides*. The model **D** calculated for the structure of cyclo- $\beta$ -tripeptide **C** (R=H). This model is in agreement with experimental facts such as very high melting points and IR spectra of cyclo- $\beta$ -tripeptides. The cyclo- $\beta$ -tripeptide **1** is deprotected using the Li-salt effect in THF, yielding the water-soluble triacid **2**. The bundle of structures **E** is representative of the *solution structure* of **2** in water, as determined by NMR spectroscopy. The substituents occupy lateral positions on the trilactam ring, a conformer with axial dispositions of the side chains (obtained by ring inversion) as in **F** is destabilized by  $A^{1,3}$  strain. For the corresponding  $\beta^2$ -peptide (see model **G**), axial substituents should be tolerated, thus metal complexation underneath the trilactam ring may be possible with this isomer.



a large excess of LiCl (up to 23 equiv.)<sup>5)</sup>. The solution thus obtained was used for the hydrogenolytic ester cleavage over Pd on charcoal (see *Scheme*). The reaction was rather slow, and the yields in the heterogeneous mixture varied between 20 and 35%<sup>6)</sup>. Nevertheless, we were able to isolate and purify by reversed phase HPLC (eluent 1% CF<sub>3</sub>CO<sub>2</sub>H in H<sub>2</sub>O/MeCN) the triacid **2** in pure form. The compound is extremely water-soluble.

**NMR Analysis of Triacid 2.** – The NMR spectrum in D<sub>2</sub>O clearly demonstrates that the predominant conformation possesses a three-fold symmetry (by the magnetic equivalence of the NH, C(β)–H, C(α)–H, and C(α)–H' protons of all three residues). By symmetry, it is, therefore, sufficient to determine the structure of the β-tripeptide **2** by assigning only three torsion angles of one of the component β-amino acids. Scalar <sup>3</sup>J-coupling constants lead directly to the torsion angles *via* the *Karplus* equation [23–26]: The distinct *J* values observed in D<sub>2</sub>O (11.7 and 2.9 Hz for C(β)H–C(α)H<sup>ax</sup> and C(β)H–C(α)H<sup>lat</sup>) clearly indicate an antiperiplanar arrangement of the C(β)H<sup>ax</sup>–C(α)H<sup>ax</sup> protons. Furthermore, the large NH,C(β)H coupling constant (9.9 Hz)<sup>7)</sup> establishes the antiperiplanar disposition of the corresponding H-atoms. These values were used as torsion-angle restraints for the structure determination by the program X-PLOR [27]. The resulting bundle **E** of conformations, which is representative of the structure in aqueous solution, is shown in the *Scheme*.

**Discussion and Conclusions.** – It is obvious that the experimentally determined structure **E** of the twelve-membered ring is essentially identical with the trilactam components in model **D**. The structure is characterized by an all-up arrangement of the C=O groups. The laterally disposed side chains bent downwards so that the carboxylic acid groups populate the half-space underneath the peptidic macrocycle. This solution structure is very similar to the solid-state structures observed for the triolides **B**. Furthermore, the *J* values measured are nearly identical to those observed for the corresponding H-atoms in the <sub>3</sub><sub>1</sub>-helix of β-peptides [28][29]. Therefore, the structure may be referred to as a 'helix without a pitch' or, conversely, the <sub>3</sub><sub>1</sub>-helix may be considered to be composed of trilactam units cut open and bent to form a left-handed single helix pitch<sup>8)</sup>.

There is both a resemblance and a fundamental difference between the structures of **2** and of enterobactin. The twelve-membered ring of enterobactin (**A**) is essentially superimposable with that of **2**, with the NH groups in **2** replaced by the O-atom in **A**. The NH–CO–Aryl substituents on the enterobactin trilactone occupy axial positions, which, in the case of the neighboring lactone O-atoms, do not lead to any A<sup>1,3</sup> repulsion. The side chains in **2** occupy lateral positions, so that the acid groups can approach each other only when bent down, which was actually observed in aqueous solution. A complexation of a trivalent ion by the three carboxylate groups of **2** would probably

<sup>5)</sup> This effect was originally discovered with α-peptides [20–22] and is now used frequently in peptide synthesis (for a review, see [17]).

<sup>6)</sup> Cf. the debenzoylation of a cyclo-β-tetrapeptide [12].

<sup>7)</sup> The NH resonance disappears after several min due to exchange with D<sub>2</sub>O.

<sup>8)</sup> Cf. the X-ray crystal structure of a linear β-tripeptide containing a nascent helix, Fig. 3 in [30].

not be possible on steric grounds; one or two additional CH<sub>2</sub> groups in the side chains, should, however, allow for a geometry in which the trilactam derivative becomes a C<sub>3</sub>-symmetrical ligand for such an ion. Inversion of the trilactam ring conformation to put the side chains in axial positions (see model **F**) is impossible because of six massive 1,5-repulsive interactions between CH<sub>2</sub> groups and carbonyl O-atoms! An axial substitution on a trilactam structure is conceivable with β<sup>2</sup>- instead of β<sup>3</sup>-amino-acid building blocks (model **G**), when only the NH H-atoms of the three amide bonds are in a 1,5 coplanar arrangement with the side chain CH<sub>2</sub> groups.

We have started work to synthesize analogues of **2** with both possible C<sub>3</sub>-symmetrical types of substitution patterns, the one bearing elongated side chains, and the corresponding β<sup>2</sup>-amino-acid-derived isomers (*cf.* **G**).

We thank Mrs. B. Brandenberg for recording the NMR spectra.

## REFERENCES

- [1] H. Drechsel, G. Jung, *J. Pept. Sci.* **1998**, *4*, 147.
- [2] L. D. Loomis, K. N. Raymond, *Inorg. Chem.* **1991**, *30*, 906.
- [3] T. B. Karpishin, T. M. Deqey, K. N. Raymond, *J. Am. Chem. Soc.* **1993**, *115*, 1842.
- [4] S. Das, U. D. Lengweiler, D. Seebach, R. N. Reusch, *Proc. Natl. Acad. Sci. U.S.A.* **1997**, *94*, 9075.
- [5] D. Seebach, M. G. Fritz, *Intl. J. Biol. Macromol.* **1999**, in press.
- [6] D. Seebach, H. M. Müller, H. M. Bürger, D. A. Plattner, *Angew. Chem.* **1992**, *104*, 443; *Angew. Chem. Int. Ed.* **1992**, *31*, 434.
- [7] D. A. Plattner, A. Brunner, M. Dobler, H.-M. Müller, W. Petter, P. Zbinden, D. Seebach, *Helv. Chim. Acta* **1993**, *76*, 2004.
- [8] A. Brunner, F. N. M. Kühnle, D. Seebach, *Helv. Chim. Acta* **1996**, *79*, 319.
- [9] D. N. J. White, C. Morrow, P. J. Cox, C. N. C. Drey, J. Lowbridge, *J. Chem. Soc., Perkin Trans. 2* **1982**, 239.
- [10] D. Seebach, J. L. Matthews, A. Meden, T. Wessels, C. Baerlocher, L. B. McCusker, *Helv. Chim. Acta* **1997**, *80*, 173.
- [11] T. D. Clark, L. K. Buehler, M. R. Ghadiri, *J. Am. Chem. Soc.* **1998**, *120*, 651.
- [12] K. Gademann, M. Ernst, D. Hoyer, D. Seebach, *Angew. Chem.* **1999**, *111*, 1302; *Angew. Chem. Int. Ed.* **1999**, *38*, 1223.
- [13] D. Seebach, M. Overhand, F. N. M. Kühnle, B. Martinoni, L. Oberer, U. Hommel, H. Widmer, *Helv. Chim. Acta* **1996**, *79*, 913.
- [14] J. L. Matthews, M. Overhand, F. N. M. Kühnle, P. E. Ciceri, D. Seebach, *Liebigs Ann. Chem.* **1997**, 1371.
- [15] J. L. Matthews, K. Gademann, B. Jaun, D. Seebach, *J. Chem. Soc., Perkin Trans. 1* **1998**, 3331.
- [16] F. Mohamadi, N. G. J. Richards, W. C. Guida, R. Liskamp, M. Lipton, C. Caufield, G. Chang, T. Hendrickson, W. C. Still, *J. Comput. Chem.* **1990**, *11*, 440.
- [17] D. Seebach, A. K. Beck, A. Studer, in 'Modern Synthetic Methods 1995', Verlag Helvetica Chimica Acta (Basel) and VCH (Weinheim), 1995.
- [18] D. Seebach, H. G. Bossler, R. Flowers, E. M. Arnett, *Helv. Chim. Acta* **1994**, *77*, 291.
- [19] T. D. Clark, J. M. Buriak, K. Kobayashi, M. P. Isler, D. E. McRee, M. R. Ghadiri, *J. Am. Chem. Soc.* **1998**, *102*, 8949.
- [20] D. Seebach, H. G. Bossler, C. Gerber, S. Y. Ko, C. W. Murtiashaw, R. Naef, S. Shoda, A. Thaler, M. Krieger, R. Wenger, *Helv. Chim. Acta* **1993**, *76*, 1564; D. Seebach, A. Thaler, A. K. Beck, *ibid.* **1989**, *72*, 857.
- [21] A. P. Thaler, Dissertation Nr. 9454, ETH-Zürich, 1991.
- [22] A. Thaler, D. Seebach, F. Cardinaux, *Helv. Chim. Acta* **1991**, *74*, 628.
- [23] M. Eberstadt, G. Gemmecker, D. F. Mierke, H. Kessler, *Angew. Chem.* **1995**, *107*, 1813; *Angew. Chem. Int. Ed.* **1995**, *34*, 1671.
- [24] K. Wüthrich, in 'NMR of Proteins and Nucleic Acids', John Wiley and Sons, New York, 1986.
- [25] A. d. Marco, M. Llinas, K. Wüthrich, *Biopolymers* **1978**, *17*, 617.
- [26] A. Pardi, M. Billeter, K. Wüthrich, *J. Mol. Biol.* **1984**, *180*, 741.
- [27] A. T. Brünger, 'X-PLOR Manual V3.0', Yale University, New Haven 1992.

- [28] D. Seebach, S. Abele, K. Gademann, G. Guichard, T. Hintermann, B. Jaun, J. L. Matthews, J. V. Schreiber, L. Oberer, U. Hommel, H. Widmer, *Helv. Chim. Acta* **1998**, *81*, 932.
- [29] K. Gademann, B. Jaun, D. Seebach, R. Perozzo, L. Scapozza, G. Folkers, *Helv. Chim. Acta* **1999**, *82*, 1.
- [30] D. Seebach, M. Overhand, F. N. M. Kühnle, B. Martinoni, L. Oberer, U. Hommel, H. Widmer, *Helv. Chim. Acta* **1996**, *79*, 913.

*Received April 22, 1999*